

The Effects of Newcastle virus Histologically Suspension on Some Organs of White Mice (Balb /c)

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Abstract

Newcastle disease is a contagious bird disease affecting wild avian species and many domestic, it may be transmissible to humans. Detection of Newcastle virus confirmed by rapid test technique (Immunochromatography). ten sample (10%) out of 100 faeces sample collected from chickens were positive. Twelve mice used in this study divided into two groups, first group consist of six mice induced with 0.2 ml from one positive sample of Newcastle virus suspension to evaluate some histopathological changes caused by Newcastle virus in small intestine and stomach of mice, second group induced with 0.2 ml from phosphate buffer saline only. The results revealed that histopathological changes in liver induced by using 0.2 ml from the positive Newcastle virus suspension as hyperplasia in Kupfer cells. Lung of mice treated with this concentration of Newcastle virus has partial pneumonia and thickening the intra alveolar walls. While brain of mice treated with same concentration suffering from increase in number of glial cells (gliosis) and apoptosis in brain cells.

Key words: Newcastle virus, histologically suspension

Introduction

Newcastle disease is caused by avian paramyxovirus serotype-1 (APMV-1), which is also disease virus (NDV). It is a highly contagious viral disease that affects domesticated and wild bird species throughout the world as well as NDV is a human pathogen and the most common sign of infection in humans is conjunctivitis that develops within hours of NDV exposure to the eye (1,2). However, disease host species and in different geographical locations. NDV is classified in the genus Avulavirus within subfamily Paramyxovirinae, family Paramyxoviridae and order Mononegavirales³. This enveloped virus has a negative sense non-segmented, single stranded RNA genome has 15198 nucleotides in length⁴. The genome encodes six structural and two non-structural proteins. Based on the fusion (F) gene strains are classified into lineages or genotypes; however the discrepancies between the two classification systems are nominal (5,6,7). NDV is spread primarily through direct contact between healthy birds and the bodily discharges of infected birds. The disease is transmitted through infected birds' droppings and secretions from the nose, mouth, and eyes. Clinical manifestation or severity of the ND depends largely upon the isolates involved in disease outbreak⁸. Based upon

pathogenicity, these strains are commonly categorized into velogenic (mesogenic and lentogenic types)⁹. The varying level of pathogenicity is attributed to amino acid sequence motif present in protease cleavage site of the precursor F protein (10,11).

Materials and Method

1-Samples collection:

A total of 100 faeces samples were collected from chicken suffering from clinical signs and symptoms of severe greenish watery diarrhea. Detection of Newcastle virus performed by rapid test (Immunochromatography) supplied from Biochek company –USA. The positive samples for Newcastle by rapid test were diluted with phosphate buffer saline or normal saline and stored at -20 °C in freeze. One positive sample of Newcastle was further used for the experimental study on laboratory animals (mice) for evaluation the effects of Newcastle disease on histological sections of these mice.

2-Experimental study : A total of 12 male mice species Balb/c have aged two month and weight 100-120 g divided into two groups, the first group consist of six mice injected orally with 0.2ml of Newcastle virus suspension for one positive sample. The other as

control group was received 0.2 ml of sterile phosphate buffer saline (PBS) according to methods of^(12,13) after 4-6 days clinical signs were recorded in infected animals . were observed Experimental mice were sacrificed after anesthetization by chloroform and open abdomen cavity by medical scissors, tissue from small intestine , stomach ,liver were collected from the experimentally infected mice and placed in formalin 10% for histopathological examination in later. Histological sections and staining were prepared according to methods described by¹⁴. The histopathological changes were observed by Dr. Nemah . H. AL-jabori /college of medicine / Babylon university under the magnification power 10X and 40 X of light microscope.

Results

Histological changes of current study observed in liver , lung and brain of mice infected with 0.2ml from newcasstle virus suspension , these changes shown in figure1,3,5, while figures 2,4,6 represented control group of mice treated with 0.2ml phosphate buffer saline. In this results figure (1) represented the liver of mice treated with 0.2ml from Newcastle virus suspension shows hyperpalsia in kuffer cells. The results in figure (3) indicated to lungs of mice treated with 0.2ml concentration from virus Newcastle suspension revealed to partial pnemonia and thicking the intra alveolar walls .While the figure (5) revealed to the brain of mice treated with 0.2ml concentration from newcasstle virus suspension revealed to increase in number of glial cells (glialosis) and apoptosis in brain cells. Finaly the figures 2,4,6 revealed to liver ,lung and brain respectively for control mice treated with phosphate buffer saline. No histological changes observed in liver ,lung and brain of control mice group

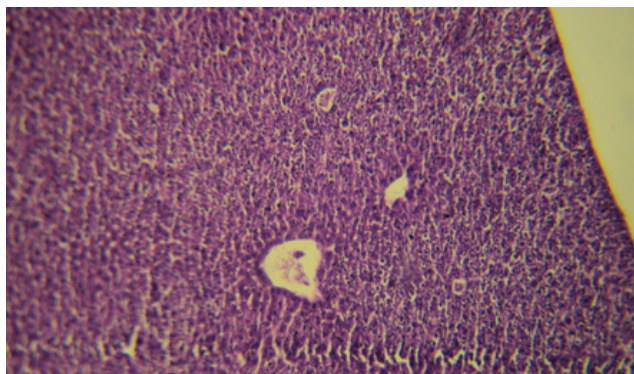


Figure (1) liver of mice treated with 0.2 ml of Newcastle virus suspension . The slide shows hyperpalsia in kuffer cells. H&E 20X

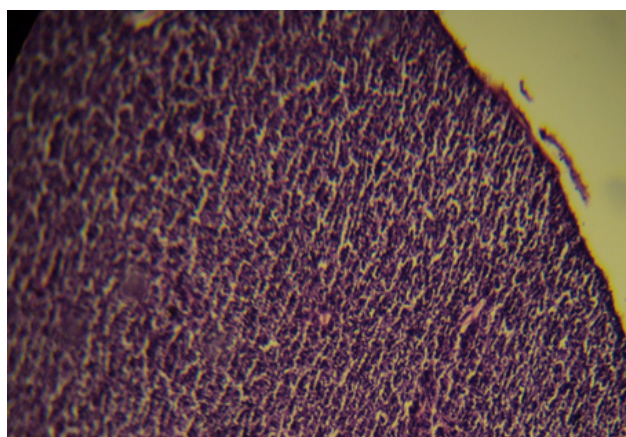


Figure (2) Control of liver mice treated with 0.2 ml phosphate buffer saline has normal hepatocytes . H&E 20X

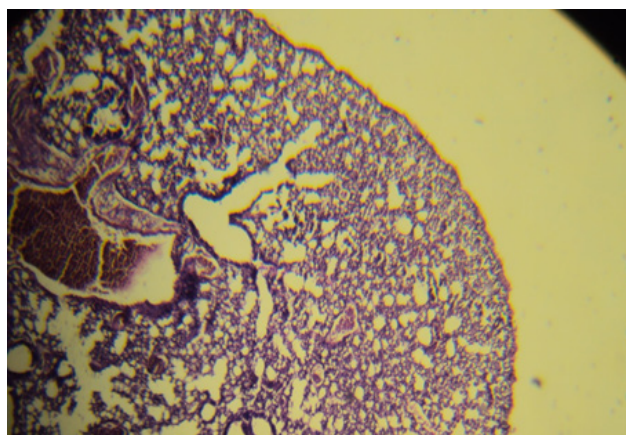


Figure (3) Lung of mice treated with 0.2 ml of Newcastle virus suspension . The slide shows partial pnemonia and thicking the intra alveolar walls. H&E20X

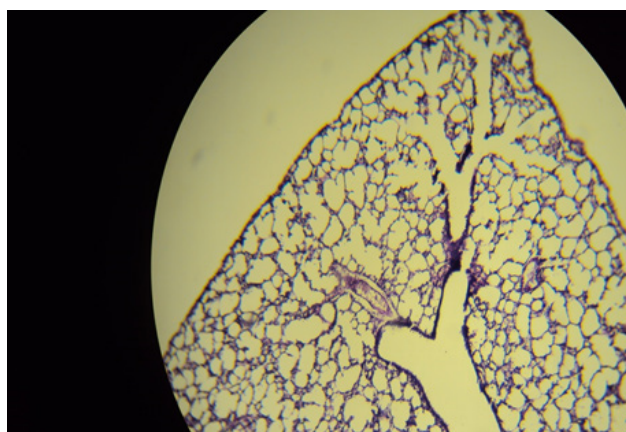


Figure (4). Control of lung mice treated with 0.2 ml phosphate buffer saline has normal intra alveolar walls . H&E 20X

Discussion

The authors consider lesions of Newcastle virus most prominent duodenum jejunum and ileum. Even in birds showing neurological signs prior to death, little evidence is found nervous system. Lesions are usually present in the respiratory tract when clinical

signs indicate involvement¹⁵. The results of present study about effects of Newcastle virus suspension on liver, lung, and brain of mice which experimentally infected revealed to histopathological changes within liver, lung and brain of mice infected with 0.2ml from Newcastle virus suspension. These histopathological changes were observed in figure 1, 3, 5. Figure (1) showed histological changes in liver represented in hyperplasia in Kupfer cells. This result is similar to recent studies mentioned that Newcastle virus outbreak in a poultry facility in Japan was characterized, among other lesions, by hepatic necrosis¹⁶.

The results of lung mice infected with same concentration of this virus revealed to partial pneumonia and thickening of the intra-alveolar walls. This result is accepted with other reported findings that some Newcastle virus strains have been shown experimentally to cause moderate lesions in the respiratory system, these changes were obtained only through aerosolization or use of very high viral titers direct air sac instillation of the virus^(17,18). While the results of lung mice infected with same concentration from Newcastle virus suspension recorded an increase in number of glial cells (gliosis) and apoptosis in brain cells. This result is in agreement with some studies mentioned finding interactions between the Newcastle disease virus and mouse tissues^(19,20). Causes of histopathological changes in liver, lung and brain of infected mice perhaps due to virulence of Newcastle virus and rapid replication effect on three systems are digestive system (e.g: liver), respiratory system (e.g: lung) and nervous system e.g: brain.^(21,22,23)

Conclusion

The Newcastle virus suspension caused clear histological changes in liver, lung and brain of mice (Balb/c).

Financial Disclosure: There is no financial disclosure.

Conflict of Interest: None to declare.

Ethical Clearance: All experimental protocols were approved under the College of biotechnology, Al-Qasim Green University, Iraq and all experiments were carried out in accordance with approved guidelines.

References

1. Swayne DE, King DJ. Avian influenza and Newcastle disease. *J. Am. Vet. Med. Assoc.*, 2003;222 (1) :1550-1534...
2. Seal BS, King DJ, Meinersmann RJ. Molecular evolution of the Newcastle disease virus matrix protein gene and phylogenetic relationships among the paramyxoviridae. *Virus Res.*, 2000;66:1-11...
3. Alexander D. Newcastle disease, other avian paramyxoviruses and pneumovirus infections. In *Diseases of Poultry*. 4th edition. Edited by Saif YM, Barnes HJ, Glisson, JR; Fadly, AM LRM and Swayne, D.: Iowa State University: 2003;63-99.
4. Kolakofsky D, Roux L, Garcin D, Ruigrok RW. Paramyxovirus mRNA editing the "rule of six" and error catastrophe: a hypothesis. *J. Gen. Virol.*, 2005;86:1877-1869.
5. Aldous EW, Mynn JK, Banks J, Alexander DJ. A molecular epidemiological study of avian paramyxovirus type 1 (Newcastle disease virus) isolates by phylogenetic analysis of a partial nucleotide sequence of the fusion protein gene. *Avian Pathol*, 2003; 32: 239-256.
6. Czegledi A, Ujvari D, Somogyi E, Wehmann E. genome size category of avian paramyxovirus serotype 1 (Newcastle disease virus) and evolutionary implications. *Virus Res.*, 2005;120:36-48.
7. Diel DG, Susta L, Cardenas G, Killian ML. Complete genome and clinicopathological characterization of a virulent Newcastle disease virus isolate from South America. *J. Clin. Microbiol.*, 2012;50:378-387..
8. Miller PJ, Decanini EL, Afonso CL. Newcastle disease: evolution of genotypes and the related diagnostic challenges. *Infect. Genet. Evol.* 2010;35:10: 26.
9. OIE: Newcastle disease. In Chapter 2115 OIE Manual of Standards for Diagnostic Tests and Vaccines, in *Manual of Diagnostic Tests and Vaccines for Terrestrial Animals: Mammals, Birds and Bees*. Paris: Office International des Epizooties, 2004;270-282..
10. Herczeg J, Wehmann E, Bragg RR, Travassos D. Two novel genetic groups (VIIb and VIII) responsible for recent Newcastle disease outbreaks in Southern Africa, one (VIIb) of which reached Southern Europe. *Arch. Virol.*, 1999 ;144:2087-2099.
11. Munir M, Cortey M, Abbas M, Qureshi ZU.

- Biological Newcastle disease virus isolated from outbreaks in commercial poultry and from backyard poultry flocks in Pakistan. *Infect. Genet. Evol.*, 2012;12:1010–1019.
12. Debouck P, Pensaert M. Experimental infection of pigs with Belgian isolates of the porcine rotavirus's. *Vet. MedB*, 1979;26:517-526.
 13. Bhriugu K, Nagendra N, Durlav P, Sudip C. Experimental infection of pigs with group rotavirus and enterotoxigenic *Escherichia coli* in India gross, Histopathological and immunopathological study, *Veterinaria Italiana*, 2011;47 (2): 117-128..
 14. Luna LG. Manual of histological staining methods of the Armed Forces Institute of Pathology, 3rd Ed. McGraw Hill. New York. 1968;195 - 196.
 15. Alexander D. Newcastle disease and other avian paramyxoviruses. *Rev. sci. Tech. Off. int. Epiz.*, 2000;19 (2): 443-462.
 16. Nakamura K, Ohtsu N, Nakamura T. Pathologic and immunohistochemical studies of Newcastle disease (ND) in broiler chickens vaccinated with ND: severe nonpurulent encephalitis and necrotizing pancreatitis. *Vet. Pathol.*, 2008;45:928–9339.
 17. Ali HA, Ahmed OH. Molecular Identification of *Entamoeba Histolytica* In Amoebiasis Patients. *J.Global Pharma Tech.* 2018; 10 (10 (Suppl.)):403-407
 18. Ahmed OH. EFFECT OF FUMONISIN B1 ON HISTOLOGY OF SPLEEN OF BROILER CHICKEN *GALLUS GALLUS*. *Biochem. Cell. Arch.* 2018;18(2): 1755-1761,
 19. Abdul-Aziz TA, Arp LH, Pathology of the trachea in turkeys exposed by aerosol to lentogenic strains of Newcastle disease virus. *Avian Dis*, 1983;27:1002–1011.
 20. Joseph S. Interactions of the Newcastle disease virus with mouse tissues. *Archiv of virology.*, 1961 ;10(1): 103-125.
 21. Ahmed OH. Histological Effect of Androgenic Anabolic Steroide Dianabol in Heart and Some Blood Parameters of Male Albino Rats. *J.Global Pharma Tech.* 2018;10(03): 215-219
 22. Ali HA, Ahmed OH. Immunohistological Study of ER, pR, and Fler2neu status in Breast carcinoma. *Indian J. Public Hralth Research & Development.* 2019;10(9).