

Immunomodulation of Iraqi *Lycium barbarum* Carotene *in vitro*

Zainab Yaseen Mohammed Hasan¹; Najwa ShAhmed¹; Raghad K. Al-Lihaibi²

¹Biotechnology Research center Al-Nahrain University, Iraq Baghdad, ²College of Pharmacy Al-Nahrain University, Iraq Baghdad

Abstract

The contents of *Lycium barbarum* active constituents as wild Iraqi plant was extracted then estimated qualitatively and quantitatively by HPLC method. Effect of the extracted carotene was employed in the current study on the normal human blood lymphocytes to estimate the extract effect on IL-6 and IL-8 levels as immune modulating agent. *L. barbarum* considered as a good source for carotene as it's contained was 0.287 mg/g dried fruits. The current study results about the effect of extracted carotene on normal human lymphocytes and interleukins 6 and 8 levels showed that the compound can enhance cell-mediated immune response through enhance lymphocytes proliferation with increasing in IL-8 and decrease IL-6 level

Keywords: *Lycium barbarum* active, lymphocytes, *L. barbarum* considered, proliferation, carotene.

Introduction

Wild Iraqi *Lycium barbarum* is rich with carotenoids that make the fruit possessed a beautiful orange color⁽¹⁾. *Lycium* was very important medicinal plant that possessed several biological activities, among them; hepatic protection from being damaged, immune boosting agent, and can reduce the harmful effects of the chemotherapy drugs and radiotherapy treatment⁽²⁾, also the regular carotene intake with daily foods as rich carotene vegetables diets might play an important role in reduction of different types of cancer⁽³⁾. Due to the potent antioxidant properties of such compounds, many researches correlated between their importance in curing several chronic diseases as well as a potent biological effect on human⁽²⁾. Moreover, the immune modulation effects of beta carotene were suggested to be a major factor in fighting cancer, by boosting the immune system⁽⁴⁾. This study projected on extraction of total carotene from wild *L. barbarum* to insure the immune boosting activity for these components and declare how it can regulate some interleukin levels of a normal human blood lymphocyte culture.

Material and Method

Extraction the total carotene from the fruit⁽⁵⁾:

Ripe *L. barbarum* fruits were dried and powdered by blender. A quantity of one gram dried fruit powdered was homogenized well with 3 ml distilled water, to be mixed then with 2 ml absolute ethanol with continuous agitation. After filtration, total carotene was extracted by 10 ml n-hexane using separator funnel. The organic hexane layer was subjected to HPLC assay.

Immunomodulation Determination (in vitro)⁽⁶⁾:

In this study about five ml blood samples were taken from peripheral vein of healthy volunteers with age of (25-35) year's old whom never using any drugs before 2 weeks. Each blood sample was suspended into 3 ml lymphocyte separation fluid (sp. gr. 1.077 g/L) in vacuumed tubes separately to be centrifuged then for half an hour at 2000 rpm. The isolated cells of lymphocytes were collected by sterile Pasteur pipette and transferred each sample into separated vacuumed tubes after washing the pellets, then suspended in with 5 ml RPMI-1640 containing 10% fetal calf serum. All tubes were incubated overnight at 37°C in 5% CO₂ incubator.

Corresponding Author:

Zainab Yaseen Mohammed Hasan

Email: Zainaby2003@yahoo.com

i-Measurement of the Viable Lymphocytes by MTT Assay⁽⁷⁾.

According to Freshney 2012 protocol, the effect of extracted carotene on normal lymphocyte culture was determined by preparing different carotene concentrations (500, 250, 125, 62.5, 31.25, 15.625, and 7.8125) $\mu\text{g/ml}$, then sterilized with disposable millipore filter. Aliquot of 100 μl lymphocyte cell culture was treated with each carotene concentrations in triplicate with control positive that employed as 0.1% PHA solution (phytohemagglutinin), and control negative represented by cells suspended in medium without any treatment. The 96 well plate was incubated for 20 hours, then centrifuged for 5 minutes at 1500 rpm. The medium was gently aspirated and MTT dye (2 mg/ml) to all wells was added. The microtiter plate was incubated again for about 3-4 hours. Crystals were formed by living lymphocytes might dissolved with 100 μl DMSO and the intensity of purple color for each well was read at 620 nm and recorded within short period by ELISA reader.

-Percentage of viable Lymphocytes can be calculated with the following equation:

$$\left[\frac{\text{Absorbance of the test}}{\text{Absorbance of negative control}} \right] \times 100.$$

Determination of the Cytokine Level by ELISA Technique⁽⁸⁾

-For this assay two tissue culture plates of 24 wells, were seeded with 1 ml of isolated lymphocytes suspended cells (1×10^6 cell/well). One plate was treated with 1 ml from the extracted carotene in three selected concentrations (100, 500, 250) $\mu\text{g/ml}$ in triplicate to be incubated for two hours, while the other plate incubated for four hours after treatment. Lymphocyte cells in growth medium alone was represented the negative control.

All wells content were pooled at the end of exposure time the end of each interval times into sterile tubes and centrifuged for 10 minutes at 2000 rpm to separate the pellets from the supernatant of each tube and kept at -20°C to be estimated by ELISA kits for IL6 and IL-8 assay.

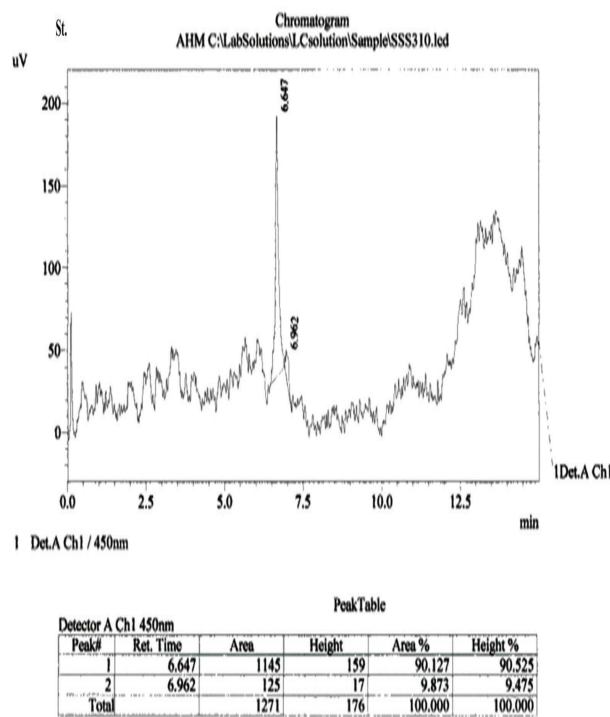
The Cytokine IL-6 and IL-8 Levels

The kit work was done according to kit protocol of United State Biological and Biochemical Reagents Trade Company specific for (IL-6 and IL-8) level, that can be calculated through plotting standard curve between concentration versus absorbance read by ELISA microplate reader (Olympus/Japan) at 450 nm were plotted, then IL-6 and IL-8 levels for each sample was calculated and then evaluated statistically.

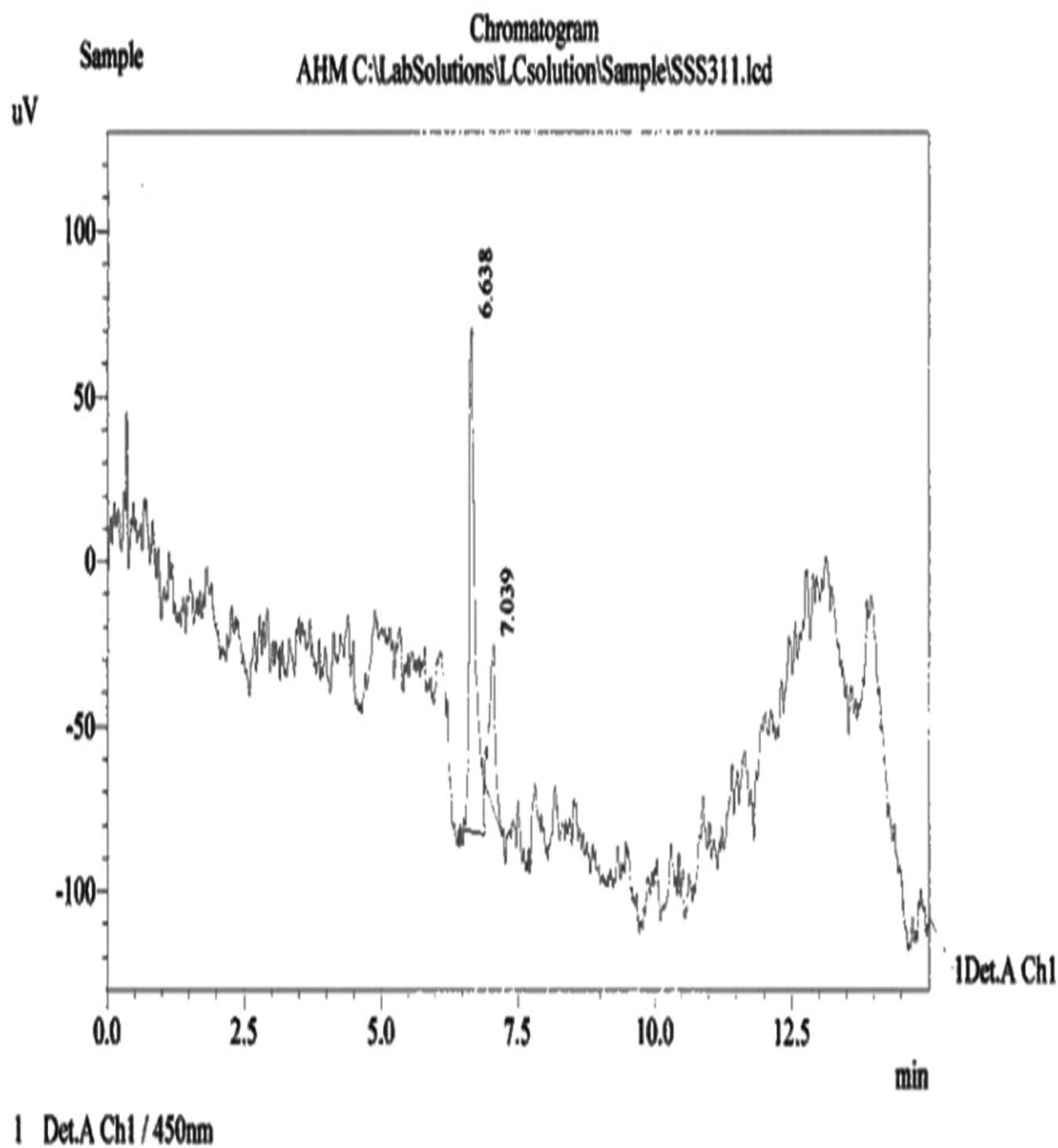
Results

Content of Total Carotene Determination by HPLC

The retention time for the β -carotene standard was (6.647 minutes) as shown in figure(1). HPLC chromatogram of the *Lycium barbarum* extracted carotene showed a presence of sharp peak with retention time (6.638) minutes, figure(2).



Figure(1) HPLC Chromatogram of the standard β -carotene



PeakTable

Detector A Ch1 450nm					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.638	1162	154	76.226	75.752
2	7.039	362	49	23.774	24.248
Total		1524	203	100.000	100.000

Figure (2) HPLC Chromatogram of the extracted carotene.

Total extracted carotene concentration can be calculated through data applied for area under the curve at retention time 6.638 minutes of the extract and 6.647 minutes of the standard with the following equation:

$$\frac{\text{Peak area of extracts}}{\text{Peak area of standard}} \times \text{Standard solution concentration} \\ \times \text{total volume of extract} \\ = \text{total carotene (mg) in 1g dried fruit powder}$$

That is

$$\frac{154}{159} \times 0.0018 \frac{\text{mg}}{\text{ml}} \times 165\text{ml} = 0.287 \text{ mg}$$

MTT Result

Lymphocyte Proliferation determination by MTT assay:

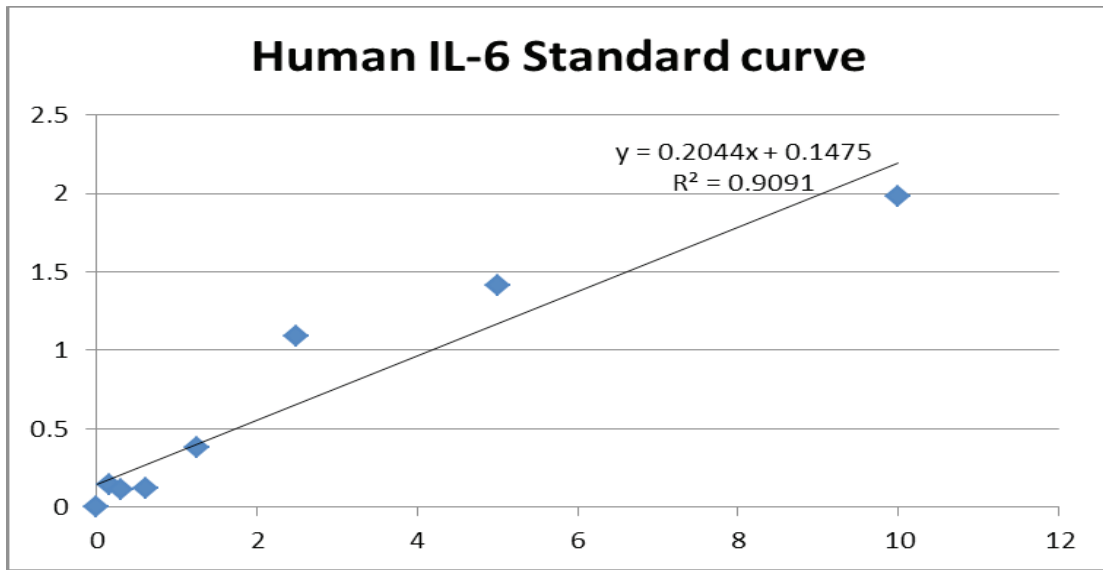
All Results were summarized in Figure(1) showed that the seven concentrations of extracted carotene affect normal human Lymphocyte by proliferation in corresponding with negative control.

Levels of IL-6

Table (1) showed the average absorption of standard solutions for IL-6 , from which a standard curve, figure (2), was plotted to get the straight line equation

Table(1) Standard IL-6 concentration and average absorption

IL-6 standard concentration(pg/ml)	Average absorption
0	0
0.156	0.144
0.312	0.112
0.625	0.117
1.25	0.38
2.5	1.093
5	1.411
10	1.978



Figure(3) Interleukin-6 standard curve

$Y=0.2044X+0.1475$

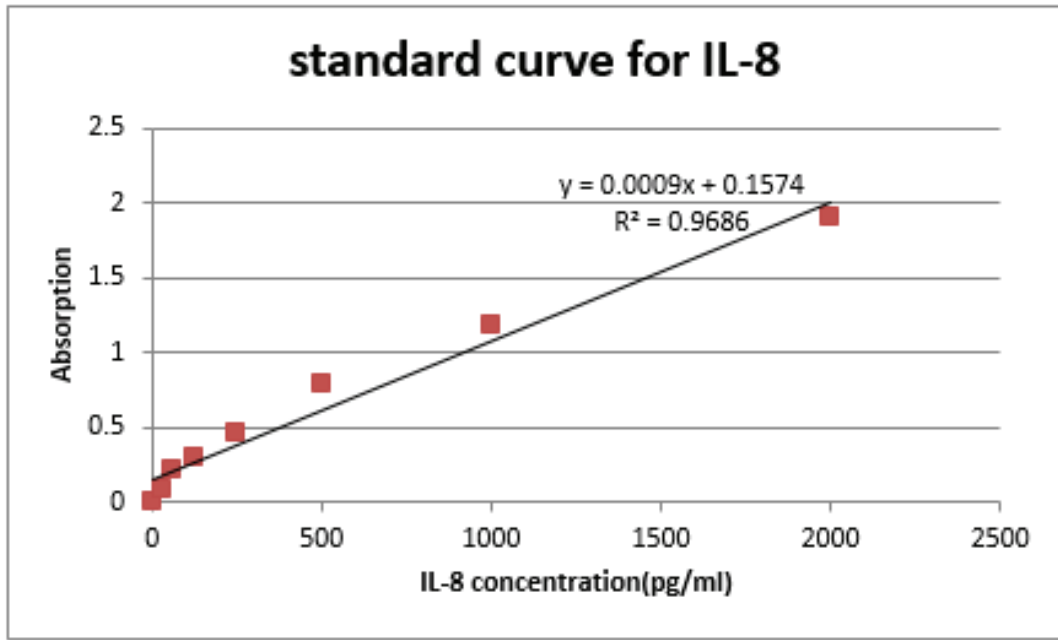
{ $X=Y-0.1475/0.2044$ pg/ml} calculation of the IL-6 level.

Levels of IL-6 from lymphocytes treated with three concentrations of extracted carotene were shown in table(2) after two intervals 2 and 4 hours of exposure.

Interleukin 6 (IL-6) encoded by the *IL6* genes; produced from different cells such as smooth cells in blood vessels. The IL-6 main action was indicated as a pro- inflammatory cytokine and myokine⁽⁹⁾, which

mediated by inhibition the secretion of TNF- α and IL-1 with activation of IL-10⁽¹⁰⁾. A decrease in IL-6 level secreted from treated lymphocytes with carotene in different concentration and 2 or 4 hours exposure, were shown in the current study. Dietary carotenoids and retinoids played an obvious roles in boosting human's innate and acquired immunity against inflammation⁽¹¹⁾. The current research could be a preliminary *invitro* study involve the beneficial effect of *L.barbarum* carotene, in spite that in a study concluded that the carotene decrease IL-6 level due to its suppression of transcription of this interleukin⁽¹²⁾.

Levels of IL-8:



Figure(4) Interleukin-8 standard curve

Figure(4) showed a standard curve for IL-8 and the straight line equation

$$Y=0.0009X+0.1574$$

{ $X=Y-0.1574/0.0009$ } calculating IL-8 level (pg/ml)

After application of IL-8 equation, the levels of treated lymphocytes with three concentrations from the extracted carotene were determined for 2 and 4 hours intervals, table(2).

Table(2) Level of IL-8 for Lymphocytes treated with *Lycium carotene*

Lyciumcarotene Extract concentration(mg/ml)	IL-8 Level(pg/ml) After 2hr.exposure	IL-8 Level(pg/ml) After 4hr. exposure
125	2922.8888	3167.333
250	2951.7777	2856.2222
500	2986.8888	3062.8888
Control	2799.000	2799.550

Results showed that IL-8 level elevated for blood lymphocytes treated with different concentrations and two intervals time of exposure. Interleukin 8 secreted by macrophages and epithelial endothelial cells, is an important mediator of the innate immune system (9).IL-8 acted as *neutrophil chemotactic factor*by induction

of chemo-taxis for neutrophils and granulocytes, in order to migrate toward infection. Moreover, it acted to stimulate phagocytosis at site of inflammation as in bronchitis and respiratory viral diseases (13). *Lyciumbarbaum* carotene play unportent role in treating different diseases via improvement of body immunity⁽¹⁴⁾.

The current results were agreed with a study by Lin and co about the effect of edible carotene on normal peripheral blood lymphocytes⁽¹⁵⁾.

Conclusion

The contents of *Lyciumbarbarium* active constituents as wild Iraqi plant was considered a good source for carotene which contained 0.287mg/g dried fruits. The current study focused on the effect of extracted carotene on normal human lymphocytes and interleukin 6 and 8 levels. The study general conclusion insured that this compound can enhance cell-mediated response for immune system. *Lyciumbarbarum* carotene possess major role in boosting human immunity that highlighted their importance as food as well as a promise medical component.

Acknowledgment: The authors are grateful to Rsearchers at the Biotechnology Research Center, (University of Al-Nahrain) for their scientific support.

Conflict of Interest: There is no conflict of interest among the authors.

Funding: Self

Ethical Clearance: This study is ethically approved by the Institutional ethical Committee.

References

- 1- Yin G, Dang Y. Optimization of extraction technology of the *Lycium barbarum* polysaccharides by Box- Behnken statistical design. Carbohydrate polymers. 2008 Nov 4;74(3):603-10.
- 2- Bungheza IR, Marius AS, Marian N, Georgeta R, Rodica-Mariana I. Obtaining of carotenoid extract from *Lycium chinense* and characterization using spectometrical analysis. Digest Journal of Nanomaterials & Biostructures (DJNB). 2012 Apr 1;7(2).
- 3- Amagase H, Farnsworth NR. A review of botanical characteristics, phytochemistry, clinical relevance in efficacy and safety of *Lycium barbarum* fruit (Goji). Food research international. 2011 Aug 1;44(7):1702-17.
- 4- Chang RC, So KF. Use of anti-aging herbal medicine, *Lycium barbarum*, against aging-associated diseases. What do we know so far?. Cellular and Molecular Neurobiology. 2008 Aug 1;28(5):643-52.
- 5- Lam KW, But P. The content of zeaxanthin in Gou Qi Zi, a potential health benefit to improve visual acuity. Food chemistry. 1999 Nov 1;67(2):173-6.
- 6- Fernandez-Botran R, Vetvicka V. Advanced methods in cellular immunology. CRC Press; 2000 May 26.
- 7- Manshoo MA, Al-Halbosiy MM, Radhwan MM. IN VITRO CYTOTOXIC ACTIVITY OF SOME WILD PLANTS EXTRACTS AGAINST RAW264. 7 CELL LINE. Plant Archives. 2019;19(2):3983-6.
- 8- De-Waal M, Moree KM. The cytokine Handbook, 3rd edn: Thompson.
- 9- Peter JD, Seamus JM, Dennis RB, Ivan MR. Roitt's Essential Immunology.
- 10- Milani A, Basirnejad M, Shahbazi S, Bolhassani A. Carotenoids: biochemistry, pharmacology and treatment. British journal of pharmacology. 2017 Jun;174(11):1290-324.
- 11- Tanaka T, Narazaki M, Kishimoto T. IL-6 in inflammation, immunity, and disease. Cold Spring Harbor perspectives in biology. 2014 Oct 1;6(10):a016295.
- 12- Katsuura S, Imamura T, Bando N, Yamanishi R. β -Carotene and β -cryptoxanthin but not lutein evoke redox and immune changes in RAW264 murine macrophages. Molecular nutrition & food research. 2009 Nov;53(11):1396-405.
- 13- Ang Z, Koean RA, Er JZ, Lee LT, Tam JK, Guo H, Ding JL. Novel AU-rich proximal UTR sequences (APS) enhance CXCL8 synthesis upon the induction of rpS6 phosphorylation. PLoS genetics. 2019 Apr 10;15(4):e1008077.
- 14- Tang L, Zhang Y, Jiang Y, Willard L, Ortiz E, Wark L, Medeiros D, Lin D. Dietary wolfberry ameliorates retinal structure abnormalities in db/db mice at the early stage of diabetes. Experimental biology and medicine. 2011 Sep;236(9):1051-63.
- 15- Bessler H, Salman H, Bergman M, Alcalay Y, Djaldetti M. In vitro effect of lycopene on cytokine production by human peripheral blood mononuclear cells. Immunological investigations. 2008 Jan 1;37(3):183-90.